

SYNTHESIS OF 5,6-*cis*-PENEMS

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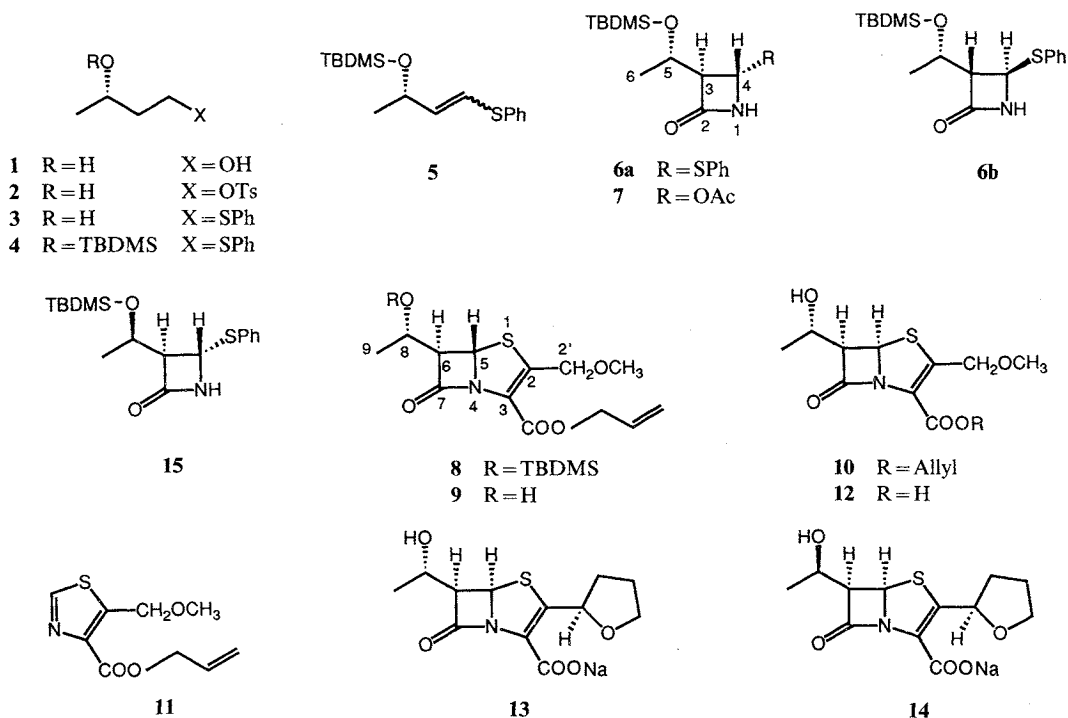
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Although the synthesis of naturally occurring 5,6-*cis*-carbapenem antibiotics such as carpetimycins¹⁾ has been extensively studied²⁾, synthesis of 5,6-*cis*-penem compounds have not been so extensively investigated. A 5,6-*cis*-penem has been obtained as a minor product in the synthesis of thermodynamically stable 5,6-*trans*-penem by intramolecular Wittig reaction³⁾, and another report described that a kinetically controlled intramolecular substitution reaction at C-4 of azetidinone⁴⁾ afforded *cis*-penem. We have observed an effective photoisomerization of (5*R*,6*S*)-5,6-*trans*-penems to thermodynamically unstable and biologically inactive (5*S*,6*S*)-5,6-*cis*-penems *via* inversion of C-5 configuration⁵⁾. This suggests that (5*R*,6*R*)-5,6-*cis*-penem derivatives can be obtained when (5*S*,6*R*)-5,6-*trans*-penem derivatives are isomerized at C-5

by photoirradiation. We herein wish to describe a synthesis of biologically active (5*R*,6*R*)-5,6-*cis*-penems applying a photoisomerization reaction to (5*S*,6*R*)-5,6-*trans*-penem derivatives which were prepared from (4*S*)-4-acetoxyazetidinone (7) derived from (*S*)-1,3-butanediol by use of the previously reported method^{6,7)}.

Starting from (*S*)-1,3-butanediol (1), (3*R*,4*S*,5*S*)-4-acetoxyazetidinone (7) was synthesized by 7 steps involving a coupling reaction of chlorosulfonyl isocyanate (CSI) with vinylsulfide (5) as key step^{6,7)}. Tosylation of butanediol (1) with tosylchloride (1.1 equiv) and lutidine (2.1 equiv) gave the monotosylate (2). This tosylate (2) was converted to phenyl sulfide (3) by treatment with sodium phenylsulfide in THF, followed by protection of the hydroxy group by the *tert*-butyldimethylsilyl (TBDMS) group. Chlorination of 4 by *N*-chlorosuccinimide (1.5 equiv) in CCl₄ at room temperature followed by dehydrochlorination by treatment with lithium carbonate (1.5 equiv) lithium iodide (1 equiv) and lithium perchlorate (1 equiv) in DMF afforded a mixture of *cis/trans* isomers (5) in 1:2.5 ratio. This mixture was then reacted with CSI (1.1 equiv) in diisopropyl ether at room temperature for 2 hours to give a mixture of diastereomeric (3*R*,4*S*,5*S*) and

Scheme 1.



(3*S*,4*R*,5*S*)-4-phenylthioazetidinone (**6a** and **6b**) in 2:1 ratio after reductive workup with benzenethiol and pyridine in acetone. Washing this mixture with *n*-hexane to remove a minor diastereoisomer (**6b**) and recrystallization from *n*-hexane gave the pure (3*R*,4*S*,5*S*)-compound (**6a**), $[\alpha]_D -63.6^\circ$ (*c* 0.332, CHCl₃). Treatment of phenylthioazetidinone (**6a**) with cupric acetate⁷⁾ in acetic acid at reflux for 1 hour provided the desired (3*R*,4*S*,5*S*)-4-acetoxyazetidinone (**7**). Overall yield of the acetoxyazetidinone (**7**) from butanediol (**1**) was about 30%.

This acetoxyazetidinone (**7**) was then converted to 5*S*,6*R*,8*S*-penem compounds by use of a conventional method^{6,8)}. Treatment of **7** with sodium methoxyacetyl thiolate in aqueous acetone, and subsequent treatment with allyl oxallyl chloride (1.1 equiv) gave the oxamate derivative which was heated with triethyl phosphite in xylene at 140°C to give (5*S*,6*R*,8*S*)-2-methoxymethyl penem (**8**) in 40% yield, which showed a characteristic coupling constant ($J=1$ Hz) between C-5 and C-6 protons for *trans*-penem structure. In this reaction, no

thermal isomerization was observed in contrast to the case of 2-alkylthiopenem compounds³⁾, suggesting an important role of the alkylthiol group attached to C-2 to assist thermal opening of the 5-membered ring. After deprotection of the alcohol by treatment with tetra-*n*-butylammonium fluoride in THF, a solution of the compound **9** in ethyl acetate (2 mM) was irradiated by use of high pressure UV lamp (Hanovia) through Pyrex filter⁵⁾ for 50 minutes to afford the isomerized 5,6-*cis*-penem (**10**) together with unchanged *trans*-penem (**9**) in 2:1 ratio in good yield and a minor amount of further degraded thiazole compound **11** (<5%)⁵⁾. Further irradiation increased the *cis/trans* ratio, but the yield was reduced because of a rapid degradation of *cis*-penem to compound **11**. Then, removal of the allyl group of 5,6-*cis*-penem (**10**) by treatment with tetrakis(triphenylphosphine) palladium⁹⁾ and sodium 2-ethylhexanoate in ethyl acetate provided the carboxylic acid (**12**). IR (ν_{\max} cm⁻¹ 3466, 1782, 1697) and UV (λ_{\max} nm 303) spectra, and FAB-MS (m/z 260, (M+H)⁺) of the compound **12**, indicat-

Table 1. *In vitro* activity of *cis* and *trans* penem derivatives.

| Organism | MIC (μ g/ml) | | | | |
|--|-------------------|------|-------|---------|-------------------------------|
| | 12 | 13 | 14 | SUN5555 | <i>trans</i> -12 ^a |
| <i>Staphylococcus aureus</i> 209P JC-1 | 1.56 | 0.78 | 3.13 | 0.10 | 0.10 |
| <i>S. aureus</i> Smith | 1.56 | 0.78 | 6.25 | 0.10 | — |
| <i>S. aureus</i> S-14 | 3.13 | 1.56 | 6.25 | 0.39 | 0.20 |
| <i>S. epidermidis</i> ATCC 14990 | 0.78 | 0.78 | 3.13 | 0.10 | — |
| <i>Bacillus subtilis</i> ATCC 6633 | 0.20 | 0.39 | 0.39 | <0.025 | 0.05 |
| <i>Micrococcus luteus</i> ATCC 9341 | 0.20 | 0.20 | 3.13 | 0.05 | 0.05 |
| <i>Escherichia coli</i> NIHJ JC-2 | 1.56 | 0.78 | 6.25 | 0.78 | 0.39 |
| <i>E. coli</i> KC-14 | 0.78 | 0.39 | 1.56 | 0.20 | 0.20 |
| <i>E. coli</i> KC-14/RGN823 | 50 | 50 | 25 | 0.20 | 0.20 |
| <i>E. coli</i> KC-14/Rms213 | 1.56 | 1.56 | 6.25 | 0.78 | 0.78 |
| <i>E. coli</i> KC-14/RGN238 | 1.56 | 1.56 | 6.25 | 0.39 | 0.39 |
| <i>Klebsiella pneumoniae</i> PCI 602 | 1.56 | 3.13 | 3.13 | 0.20 | 0.39 |
| <i>K. pneumoniae</i> KC-1 | 1.56 | 0.78 | 6.25 | 0.39 | — |
| <i>Serratia marcescens</i> IAM 1136 | 25 | 6.25 | 25 | 3.13 | 1.56 |
| <i>S. marcescens</i> GN 10857 | >50 | >100 | >100 | 50 | 25 |
| <i>Proteus mirabilis</i> GN 79 | 6.25 | 6.25 | 25 | 3.13 | 0.78 |
| <i>P. vulgaris</i> GN 7919 | >50 | 50 | 6.25 | 0.78 | 0.39 |
| <i>P. vulgaris</i> GN 76 | 50 | 25 | 25 | 6.25 | 3.13 |
| <i>Morganella morganii</i> IFO 3848 | 6.25 | 1.56 | 12.50 | 0.78 | 1.56 |
| <i>Providencia rettgeri</i> GN 624 | >50 | 25 | 50 | 3.13 | — |
| <i>Salmonella typhi</i> O 901 | 0.78 | 0.78 | 3.13 | 0.10 | 0.20 |
| <i>Citrobacter freundii</i> GN 7391 | >50 | >100 | >100 | 25 | 6.25 |
| <i>Enterobacter cloacae</i> 91 | 12.50 | 6.25 | 6.25 | 0.39 | 0.39 |
| <i>Pseudomonas aeruginosa</i> No. 12 | >50 | >100 | >100 | >100 | >100 |
| <i>P. aeruginosa</i> PAO-1 | >50 | >100 | >100 | >100 | — |
| <i>Acinetobacter calcoaceticus</i> AC 54 | 6.25 | 6.25 | 100 | 12.50 | 12.50 |

Medium: Sensitivity disk agar, inoculum size: 10⁶ cfu/ml.

^a (5*R*,6*S*,8*R*)-5,6-*trans* isomer of **12**.

ed penem structure, and its characteristic coupling constant ($J=4\text{ Hz}$) between C-5 and C-6 protons in its ^1H NMR and the positive Cotton effect ($\lambda_{\text{max}}\text{ nm } 250, \theta=1.75 \times 10^5$) in the CD spectrum⁵⁾ confirmed its $5R,6R$ configuration.

As described above, this synthetic route offers a new and convenient approach to the synthesis of *cis*-penem derivatives. Thus, we could synthesize the other derivatives such as **13** and **14**, the latter of which was derived from (3*R*,4*S*,5*R*)-4-phenylthioazetidinone (**15**)⁶⁾ in similar fashion.

Penem derivatives (**12**, **13** and **14**) showed good activities against various Gram-positive and Gram-negative bacteria. However, these were less active than the corresponding (5*R*,6*S*,8*R*)-5,6-*trans* isomers⁶⁾ as shown in the Table 1.

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